No. 3 Biofilter

Air Emissions Test Report at Decorative Panels International, Inc. Alpena, Michigan

Renewable Operating Permit MI-ROP-B1476-2009a SRN: B1476

Prepared for

Decorative Panels International, Inc. 416 Ford Avenue Alpena, Michigan

Bureau Veritas Project No. 11014-000010.00 March 6, 2014

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MICHIGAN DEPARTMENT OF ENVIRONMENTAL QUALITY AIR QUALITY DIVISION

RENEWABLE OPERATING PERMIT

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Reporting period (provide inclusive dates): From na To na	
Emissions test report to evaluate compliance of the No. 3 Biofilter.	
This form shall certify that the testing was conducted in accordance with the	
approved test plan and that the facility operating conditions were in compliance with	
permit requirements.	

I certify that, based on information and belief formed after reasonable inquiry, the statements and information in this report and the supporting enclosures are true, accurate and complete, and that any observed, documented or known instances of noncompliance have been reported as deviations, including situations where a different or no monitoring method is specified by the RO Permit.

Tim Clark	President	419-720-0957
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h	P. Chu	3/6/14
Signature of Responsible Official		Date



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Executive Summary

Decorative Panels International, Inc. retained Bureau Veritas North America, Inc. to test air emissions from the No. 3 Biofilter source at the hardwood manufacturing facility in Alpena, Michigan. The No. 3 Biofilter controls emissions from the EU3PRESS-AREA and is included in the FGPRESSES flexible group. The objective of the testing was to evaluate compliance of the No. 3 Biofilter source with emission limits and requirements in:

- Michigan Department of Environmental Quality (MDEQ) Renewable Operating Permit (ROP) MI-ROP-B1476-2009a for these FGMACTDDDD sources.
- 40 CFR 63, Subpart DDDD, "National Emission Standards for Hazardous Air Pollutants: Plywood and Composite Wood Products."

Compliance with the FGMACTDDDD total hazardous air pollutant (HAP) permit limits, based on the use of an add-on control device, can be demonstrated by one of the following:

- 1. 90% reduction of total HAP mass emission rate, measured as total hydrocarbons (THC), as carbon.
- 2. Total HAP concentration less than 20 ppmvd, measured as THC (as carbon).
- 3. Total HAP reduction so that methanol mass emission rate is reduced by 90%.
- 4. Total HAP reduction so that methanol concentration is less than 1 ppmvd, if the uncontrolled methanol concentration entering the control device is greater than 10 ppmvd.
- 5. Total HAP reduction so that formaldehyde mass emission rate is reduced by 90%.
- 6. Total HAP reduction so that formaldehyde concentration is less than 1 ppmvd, if the uncontrolled formaldehyde entering the control device is greater than 10 ppmvd.

Bureau Veritas measured THC, methanol, and formaldehyde at the inlet and outlet of the No. 3 Biofilter control device.

Particulate matter emissions were measured at the No. 3 Biofilter exhaust upstream of the stack's discharge to the atmosphere:

- FGPRESSES Board Press 3 and its associated board cooler air emissions controlled by the No. 3 Biofilter. Particulate matter emissions were measured at the following exhaust stack:
 - SV#3PRESS-STK68



Executive Summary

The particulate matter permit limits for the No. 3 Biofilter source are presented below:

Source Stack Parameter		Emission Limits					
FGPRESSES		Board Press 3					
SV#3PRESS-STK68	Particulate matter	0.10 lb per 1,000 lb exhaust gases on a dry basis 29.3 lb per hour					

No. 3 Biofilter ROP Emission Limits

Three, 60-minute test runs were performed under maximum routine operating conditions following United States Environmental Protection Agency (USEPA) Methods 1, 2, 3, 3A, 4, 5, 25A, 205, and 320.

Detailed results are presented in Tables 1 and 2 after the Tables Tab of this report. The following tables summarize the results of the testing conducted on January 9, 2014 compared to the permit emission limits.

No. 3 Biofilter Formaldehyde, Methanol, and THC Results

No. 3 Biofilter Formald	lehyde, Methanol, and THC	1
Destruction I	Efficiency Results	

Parameter	Units	Run 1	Run 2	Run 3	Average
Formaldehyde Destruction Efficiency	%	97.6	97.5	97.3	97.5
Methanol Destruction Efficiency	%	93.8	96.1	94.4	94.7
THC Destruction Efficiency	%	75.4	76.4	73.4	75.0

Note: Biofilter bed temperature during all three runs was 82 °F.



Executive Summary

The results of the January 9, 2014 emissions testing established the following:

• Compliance of the No. 3 Biofilter source with the formaldehyde destruction efficiency limit of 90% or greater at a biofilter bed temperature of 82 °F.

No. 3 Biofilter Particulate Matter Test Results

				Testing Results			Emission
Source Stack	Unit	Parameter	Run 1	Run 2	Run 3	Average Result	Limit
SV#3PRESS-	lb/1,000 lb	Particulate	0.030	0.0082	0.054	0.031	0.10
STK68	lb/hr	matter	7.4	2.0	13	7.4	29.3

No. 3 Biofilter Particulate Matter Results

The average results of the particulate matter emissions testing indicate the No. 3 Biofilter complies with the applicable permit limits of 0.10 pound of particulate matter per 1,000 pounds of exhaust gases on a dry basis and 29.3 pound per hour (lb/hr).



1.0 Introduction

1.1 Summary of Test Program

Decorative Panels International, Inc. retained Bureau Veritas North America, Inc. to test air emissions from the No. 3 Biofilter source at the hardwood manufacturing facility in Alpena, Michigan. The No. 3 Biofilter control emissions from the EU3PRESS-AREA and is included in the FGPRESSES flexible group. The objective of the testing was to evaluate compliance of the No. 3 Biofilter source with emission limits and requirements in:

- Michigan Department of Environmental Quality (MDEQ) Renewable Operating Permit (ROP) MI-ROP-B1476-2009a for these FGMACTDDDD sources.
- 40 CFR 63, Subpart DDDD, "National Emission Standards for Hazardous Air Pollutants: Plywood and Composite Wood Products."

Compliance with the FGMACTDDDD total hazardous air pollutant (HAP) permit limits, based on the use of an add-on control device, can be demonstrated by one of the following:

- 1. 90% reduction of total HAP mass emission rate, measured as total hydrocarbons (THC), as carbon.
- 2. Total HAP concentration less than 20 ppmvd, measured as THC (as carbon).
- 3. Total HAP reduction so that methanol mass emission rate is reduced by 90%.
- 4. Total HAP reduction so that methanol concentration is less than 1 ppmvd, if the uncontrolled methanol concentration entering the control device is greater than 10 ppmvd.
- 5. Total HAP reduction so that formaldehyde mass emission rate is reduced by 90%.
- 6. Total HAP reduction so that formaldehyde concentration is less than 1 ppmvd, if the uncontrolled formaldehyde entering the control device is greater than 10 ppmvd.

Bureau Veritas measured THC, methanol, and formaldehyde at the inlet and outlet of No. 3 Biofilter sampling location.



Particulate matter emissions were measured at the No. 3 Biofilter exhaust upstream of the stack's discharge to the atmosphere:

- FGPRESSES Board Press 3 and its associated board cooler air emissions controlled by the No. 3 Biofilter. Particulate matter emissions were measured at the following exhaust stack:
 - SV#3PRESS-STK68

Three, 60-minute test runs were performed under maximum routine operating conditions following United States Environmental Protection Agency (USEPA) Methods 1, 2, 3, 3A, 4, 5, 25A, 205, and 320. The table on the following page outlines the test methods for the test parameters, including ancillary measurements required by the USEPA methods (i.e., traverse point selection, velocity, molecular weight, and moisture content).



Table 1-1Emission Test Parameters

	So	ource	USEPA Reference		
Parameter	Inlet of No. 3 Biofilter	Outlet of No. 3 Biofilter	Method	Title	
Sampling ports and traverse points	•	•	1	Sample and Velocity Traverses for Stationary Sources	
Velocity and flowrate	•	•	2	Determination of Stack Gas Velocity and Volumetric Flow Rate (Type S Pitot Tube)	
Molecular weight	•	•	3	Gas Analysis for the Determination of Dry Molecular Weight	
Oxygen and carbon dioxide		•	3A	Determination of Oxygen and Carbon Dioxide Concentrations in Emissions from Stationary Sources (Instrument Analyzer Procedure)	
Moisture content	•	•	4	Determination of Moisture Content in Stack Gases	
Particulate matter		•	5	Determination of Particulate Matter Emissions from Stationary Sources	
Total hydrocarbons	•	•	25A	Determination of Total Gaseous Organic Concentration using a Flame Ionization Analyzer	
Gas Dilution Calibration	•	•	205	Verification of Gas Dilution Systems for Field Instrument Calibrations	
Formaldehyde and methanol	•	•	320	Measurement of Vapor Phase Organic and Inorganic Emissions by Extractive Fourier Transform Infrared (FTIR) Spectroscopy	



1.2 Key Personnel

The key personnel involved in this test program are listed in Table 1-2. Mr. Thomas Schmelter, Senior Project Manager with Bureau Veritas led the emission testing. Decorative Panels International, Inc. personnel provided process coordination and recorded operating parameters. Portions of the testing were witnessed by Messrs. Rob Dickman and William Rogers Jr., both with MDEQ.

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Table 1-2Key Personnel



2.0 Source and Sampling Locations

2.1 **Process Description**

Decorative Panels International, Inc. produces a variety of hardboard products including wall paneling, pegboard, and marker board. Hardwood chips such as aspen, ash, maple, and beech are purchased and stored in an outdoor raw material storage area and then reclaimed into silos. The wood chips are cooked and softened in one of four digesters using steam injection and then ground into wood pulp fibers.

The pulp fibers are conveyed to a forming machine, which forms a mat of unpressed hardboard. The mats are processed through a Coe dryer and are cut using a trimmer and panel brush. The mats are conveyed to one of two hardboard lines, Line 1 or 3. Line 2 was historically operated but has since been decommissioned.

On the hardboard lines, the mats enter a predryer, a press, cooler, and tempering area. The predryer ensures the mat has the desired moisture content before the mat enters presses that apply pressure and heat to form hardboard. The hardboard is coated with linseed or Oxi-Cure® oil in the tempering area. The oil tempers the board thereby increasing its strength and "paintability." Once the board has been tempered, it is superheated to cure the binding resins in the bake ovens (No. 3 Press only). The hardboard is humidified to approximate atmospheric conditions to limit warping. The boards are inspected, graded, cut, and packed for shipping.

The No. 3 Biofilter controls emissions from the No. 3 Board Press and cooler.

2.2 **Process Operating Parameters**

The process was operated under maximum routine operating conditions during testing. Table 2-1 summarizes the average board production rate and estimated capacity in thousands of square feet per hour (msfh) and day (msfd) for the No. 3 lines (EU3PRESS-AREA). Refer to Appendix E for process data recorded during testing.

Source	Production During Testing	Capacity
EU3PRESS-AREA	3/16 inch board - 14.1 msfh	290 to 310 msfd

Table 2-1Capacity of No. 3 Line

msfh: thousand square feet per hour msfd: thousand square feet per day



2.3 Control Equipment

Gaseous emissions from the No. 3 Board Press are controlled by a humidifier and Envirogen manufactured biofilter (No. 3 Biofilter). Emissions from the No. 3 Board Press enters the top of the scrubber and flows downwards in the vessel, where water treated with sodium hydroxide to maintain a neutral pH, is sprayed to humidify the inlet air to the biofilter.

As the gas mixes with the water, particulates and other pollutants are removed. The water drains to the bottom of the vessel and a portion is recirculated into the system with the remaining portion discharged to the onsite water treatment system. The flue gas exits the top of the scrubber and is directed into the No. 3 Biofilter.

The No. 3 Biofilter consists of four compartments. The air exiting the humidifier can be further humidified and heated if the facility adds steam to the ductwork upstream of the biobed compartments. The compartments contain water sprayers to maintain a moist environment, and layers of Douglas-fir bark from the western United States. The Douglas-fir bark provides an environment where biologically active microbes can oxidize and remove contaminants.

After passing through the bark the flue gas is drawn into fans that discharge the gas through stack, SV#3PRESS-STK68.

The biofilter bed temperature is continuously monitored by thermocouples in each chamber. These temperatures are reduced to 15-minute and 1-hour averages and were recorded during testing. The No. 3 Biofilter average bed temperatures during testing are presented in Table 2-2.

Test Date	Test	Bed Temperature (°F)
January 9, 2014	1	82
January 9, 2014	2	82
January 9, 2014	3	82

Table 2-2No. 3 Biofilter Bed Temperature During Testing

Refer to Appendix E for facility operating data.

2.4 Flue Gas Sampling Locations

The figures on the following pages provide photographs that show the sampling ports at the sampling locations for the No. 3 Biofilter. Appendix Figures 1 and 2 present the No. 3 Biofilter inlet and outlet sampling ports and traverse point locations.



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Figure 2-1. No. 3 Biofilter Inlet and Outlet Sampling Locations



2.5 Process Sampling Locations

Process sampling was not required during this test program. A process sample is a sample that is analyzed for operational parameters, such as calorific value of a fuel (e.g., natural gas, coal), organic compound content (e.g., paint coatings), or composition (e.g., polymers).



3.0 Summary and Discussion of Results

3.1 Objective

The objective of the testing was to evaluate compliance of the No. 3 Biofilter source with emission limits and requirements in:

- MDEQ ROP: MI-ROP-B1476-2009a for these FGMACTDDDD sources.
- 40 CFR 63, Subpart DDDD, "National Emission Standards for Hazardous Air Pollutants: Plywood and Composite Wood Products."

Compliance with the FGMACTDDDD total HAP permit limits, based on the use of an add-on control device, can be demonstrated by one of the following:

- 1. 90% reduction of total HAP mass emission rate, measured as THC, as carbon.
- 2. Total HAP concentration less than 20 ppmvd, measured as THC (as carbon).
- 3. Total HAP reduction so that methanol mass emission rate is reduced by 90%.
- 4. Total HAP reduction so that methanol concentration is less than 1 ppmvd, if the uncontrolled methanol concentration entering the control device is greater than 10 ppmvd.
- 5. Total HAP reduction so that formaldehyde mass emission rate is reduced by 90%.
- 6. Total HAP reduction so that formaldehyde concentration is less than 1 ppmvd, if the uncontrolled formaldehyde entering the control device is greater than 10 ppmvd.

Bureau Veritas measured THC, methanol, and formaldehyde at the No. 3 Biofilter sampling location. Previous testing by Bureau Veritas demonstrated compliance using Option 5 at the No. 3 Biofilter source.

Particulate matter emissions were measured at the No. 3 Biofilter exhaust upstream of the stack's discharge to the atmosphere:

- FGPRESSES Board Press 3 and its associated board cooler air emissions controlled by the No. 3 Biofilter. Particulate matter emissions were measured at the following exhaust stack:
 - SV#3PRESS-STK68



The particulate matter permit limits for the No. 3 Biofilter source are presented in Table 1-1.

Table 3-1No. 3 Biofilter ROP Emission Limits

Source Stack	Parameter	Emission Limits
FGPRESSES		Board Press 3
SV#3PRESS-STK68	Particulate matter	0.10 lb per 1,000 lb exhaust gases on a dry basis 29.3 lb per hour

3.2 Test Matrix

Table 3-2 presents the sampling and analytical matrix.

Table 3-2Test Matrix

Source	Date 2014	Run	Start Time	End Time	EPA Method
No. 3 Biofilter	Jan 9	1	10:45	15:57	
		2	16:40	17:45	1 through 5, 25A, 205, 320
		3	18:21	19:26	

3.3 Field Test Changes and Issues

The testing was performed in accordance with USEPA procedures during maximum routine operating conditions as outlined in the Intent-to-Test Plan submitted to MDEQ on April 17, 2013, and approved on May 2, 2013. The following sections describe the testing issues encountered in the field.

3.3.1 May 2013 Biofilter Reschedule

The original compliance test notification included testing the No. 3 Biofilter for MACT requirements and for particulate matter during the May 2013 testing event; however, this testing was postponed because issues were identified with the biofilters during the May 2013 testing. Therefore, the No. 3 Biofilter testing was rescheduled and completed on January 9, 2014.



3.3.2 No. 1 Biofilter Sampling Locations

Although air emissions were proposed to be measured from the No. 1 Biofilter within the Intent to Test Plan; the testing was not conducted in January 2014 due to safety concerns. The No. 1 Biofilter inlet and outlet sampling ports are positioned in the vertical and horizontal planes of the horizontal ductwork. Sampling from the vertical ports presented a safety hazard as there are no harness secure points above the work surface, the ports are elevated approximately 15 feet above the roof level, and the steel ducts were snow and ice covered. Decorative Panels will provide appropriate access at these sampling locations and the testing will be rescheduled.

3.4 Summary of Results

The results of the testing compared to permit limits are presented in Tables 3-3 and 3-4.

No. 3 Biofilter Formaldehyde, Methanol, and THC Results

Destruction Efficiency Results							
Parameter Units Run 1 Run 2 Run 3 Average							
Formaldehyde Destruction Efficiency	%	97.6	97.5	97.3	97.5		
Methanol Destruction Efficiency	%	93.8	96.1	94.4	94.7		
THC Destruction Efficiency	%	75.4	76.4	73.4	75.0		

Table 3-3No. 3 Biofilter Formaldehyde, Methanol, and THCDestruction Efficiency Results

Note: Biofilter bed temperature during all three runs was 82 °F.

The results of the January 9, 2014 emissions testing established the following:

• Compliance of the No. 3 Biofilter source with the formaldehyde destruction efficiency limit of 90% or greater at a biofilter bed temperature of 82 °F.



No. 3 Biofilter Particulate Matter Test Results

Table 3-4No. 3 Biofilter Particulate Matter Results

			Testing Results				Emigrica
Source Stack	ource Stack Unit		Run 1	Run 2	Run 3	Average Result	Limit
SV#3PRESS-	1b/1,000 lb	Particulate	0.030	0.0082	0.054	0.031	0.10
STK68	lb/hr	matter	7.4	2.0	13	7.4	29.3

The average results of the particulate matter emissions testing indicate the No. 3 Biofilter complies with the applicable permit limits of 0.10 pound of particulate matter per 1,000 pounds of exhaust gases on a dry basis and 29.3 pound per hour (lb/hr).

Detailed results are presented in the Appendix Tables 1 and 2 after the Tables Tab of this report. Graphs of the formaldehyde, methanol, and THC concentrations are presented after the Graphs Tab of this report. Sample calculations are presented in Appendix B.



4.0 Sampling and Analytical Procedures

Bureau Veritas measured emissions in accordance with procedures specified in USEPA's Standards of Performance for New Stationary Sources. The sampling and analytical methods used during this test program are listed in Table 4-1.

Method	Parameter	Analysis
EPA 1 and 2	Gas stream volumetric flowrate	Field measurement, S-type Pitot tube
EPA 3 and 3A	Oxygen, carbon dioxide, molecular weight	Fyrite® chemical absorption and paramagnetic gas analyzers
EPA 4	Moisture content	Gravimetric
EPA 5	Particulate matter	Gravimetric
EPA 25A	Total hydrocarbons	Flame ionization detector
EPA 205	Calibration gas dilutions	Field instrument verification
EPA 320	Formaldehyde and methanol	Extractive Fourier transform infrared spectroscopy (FTIR)

Table 4-1Emission Test Methods

4.1 Emission Test Methods

4.1.1 Volumetric Flowrate (USEPA Methods 1 and 2)

Method 1, "Sample and Velocity Traverses for Stationary Sources," from the Code of Federal Regulations, Title 40, Part 60 (40 CFR 60), Appendix A, was used to evaluate the sampling location, the number of traverse points for sampling, and the measurement of velocity profiles. Details of the sampling location and number of velocity traverse points are presented in Table 4-2.



Source	Sampling Location	Duct Diameter (inches)	Distance from Ports to Upstream Flow Disturbance (diameters)	Distance from Ports to Downstream Flow Disturbance (diameters)	Number of Ports Used	Traverse Points per Port	Total Traverse Points
No. 3 Biofilter	Inlet	51.0	2.6	1.5	2	12	24
No. 3 Biofilter	Outlet	51.25	5.9	3.5	2	12	24

Table 4-2Sampling Location and Number of Traverse Points

Figures 2-1 and 2-2 depict the sampling locations at the No. 3 Biofilter. Appendix Figures 1 and 2 present the No. 3 Biofilter inlet and outlet sampling ports and traverse point locations.

Method 2, "Determination of Stack Gas Velocity and Volumetric Flow Rate (Type S Pitot Tube)," was used to measure flue gas velocity and calculate volumetric flowrate. S-type Pitot tubes and thermocouple assemblies, calibrated in accordance with Method 2, Section 10.0, were used during testing. Because the dimensions of the Pitot tubes met the requirements outlined in Method 2, Section 10.1, and were within the specified limits, the baseline Pitot tube coefficient of 0.84 (dimensionless) was assigned. Refer to Appendix A for the Pitot tube inspection sheets.

Field data sheets are included in Appendix C. Computer-generated field data sheets are included in Appendix D.

4.1.2 Molecular Weight (USEPA Method 3)

Molecular weight at the No. 3 Biofilter inlet location was measured using Method 3, "Gas Analysis for the Determination of Dry Molecular Weight." Flue gas was extracted from the stack through a probe positioned near the centroid of the duct and directed into a Fyrite® gas analyzer. The concentrations of carbon dioxide (CO₂) and oxygen (O₂) were measured by chemical absorption to within $\pm 0.5\%$. The average CO₂ and O₂ results of the grab samples were used to calculate molecular weight.

4.1.3 Oxygen Content (USEPA Method 3A)

At the No. 3 Biofilter outlet location, the flue gas oxygen content were measured in order to correct the particulate matter concentrations to units of lb PM/1,000 lb of exhaust gas on a dry basis. USEPA Method 3A, "Determination of Oxygen and Carbon Dioxide Concentrations in Emissions from Stationary Sources (Instrument Analyzer Procedure)," was used to measure the oxygen concentration of the flue gas. Flue gas was extracted from the stack through:



- A stainless-steel probe.
- Heated Teflon sample line to prevent condensation.
- A chilled Teflon impinger train with peristaltic pump to remove moisture from the sampled gas stream prior to entering the analyzer.
- A Teledyne paramagnetic oxygen gas analyzer.

Data was recorded at 1-second intervals on a computer equipped with data acquisition software. Recorded O_2 concentrations were averaged over the duration of each 60-minute test run.

Prior to testing at each sampling location, a 3-point stratification test was conducted at 17, 50, and 83 percent of the stack diameter for at least twice the response time to determine the number of sampling traverse points. Since the gas stream was considered to be unstratified, a single sampling point, located near the centroid of the duct was used.

A calibration error check was performed by introducing zero-, mid-, and high-level calibration gases directly into the analyzer. The calibration error check is performed to evaluate that the analyzer respond to within $\pm 2\%$ of the calibration span. Prior to each test run, a system-bias test was performed where known concentrations of calibration gases are introduced at the probe tip to measure if the analyzers response is within $\pm 5\%$ of the calibration span. At the conclusion of the each test run, an additional system-bias check was performed to evaluate the percent drift from pre- and post-test system-bias checks. A valid system-bias check demonstrates the analyzer did not drift greater than $\pm 3\%$ of the calibration span throughout a test run.

Calibration data, along with the USEPA Protocol 1 certification sheets for the calibration gases used is included in Appendix A. Figure 3 in the Appendix depicts the USEPA Method 3A sampling train.

4.1.4 Moisture Content (USEPA Method 4)

Before testing, moisture content was estimated using previous test data, psychrometric charts, and/or saturation vapor pressure tables. This estimate was used in conjunction with preliminary velocity head and temperature data to (1) calculate flue gas velocity, 2) ideal nozzle diameter, and (3) establish isokinetic sampling rates.

At the exhaust to atmosphere sampling location, the moisture content of the flue gas was measured using the reference method outlined in Section 2 of Method 4, "Determination of Moisture Content in Stack Gases" in conjunction with USEPA Method 5 sampling train.



4.1.5 Particulate Matter (USEPA Method 5)

USEPA Method 5, "Determination of Particulate Emissions from Stationary Sources," was used to measure the filterable "front-half" particulate matter emissions. The "front half" refers to the filterable particulate mass collected from the nozzle, probe, and filter. Triplicate 60-minute test runs were performed at the outlet of the No. 3 Biofilter. Bureau Veritas' modular isokinetic stack sampling system consists of the following:

- A stainless steel or glass button-hook nozzle.
- A heated (248±25°F) stainless steel or glass-lined probe.
- A desiccated and pre-weighed 110-millimeter-diameter glass fiber filter (manufactured to at least 99.95% efficiency (<0.05 % penetration) for 0.3-micron dioctyl phthalate smoke particles) in a heated (248±25°F) filter box.
- A set of four pre-cleaned Greenburg-Smith (GS) impingers with the configuration shown in Table 4-3.
- A sample line.
- An Environmental Supply® control case equipped with a pump, dry-gas meter, and calibrated orifice.

Impinger Order (Upstream to Downstream)	Impinger Type	Impinger Contents	Amount of Contents
1	Modified	Water	100 grams
2	Greenburg Smith	Water	100 grams
3	Modified	Empty	0 grams_
4	Modified	Silica desiccant	~300 grams

Table 4-3Method 5 Impinger Configuration

Before testing, a preliminary velocity traverse was performed and a nozzle size was calculated that would allow isokinetic sampling at an average rate of 0.75 cubic feet per minute. Bureau Veritas selected a pre-cleaned stainless steel nozzle that had an inner diameter that approximates the calculated ideal value. The nozzle was measured with calipers across three cross-sectional chords to evaluate the inside diameter; rinsed and brushed with acetone; and connected to the stainless steel-lined sample probe.

The impact and static pressure openings of the Pitot tube were leak-checked at or above a velocity head of three inches of water for more than 15 seconds. The sampling train was leak-



checked by capping the nozzle tip and applying a vacuum of approximately 15 inches of mercury to the sampling train. The dry-gas meter was then monitored for approximately 1 minute to measure that the sample train leak rate was less than 0.02 cubic feet per minute (cfm). The sample probe was inserted into the sampling port to begin sampling.

Ice was placed around the impingers and the probe and filter temperatures were allowed to stabilize at 248±25 °F before each sample run. After the desired operating conditions were coordinated with the facility, testing was initiated.

Stack parameters (e.g., flue velocity, temperature) were monitored to establish the isokinetic sampling rate within ± 10 % for the duration of the test. Data were recorded at each of the traverse points.

At the conclusion of a test run and the post-test leak check, the sampling train was disassembled and the impingers and filter were transported to the recovery area. The filter was recovered using tweezers and placed in a Petri dish. The Petri dish was immediately labeled and sealed with Teflon tape. The nozzle, probe, and the front half of the filter holder assembly were brushed and, at a minimum, triple-rinsed with acetone to recover particulate matter. The acetone rinses were collected in pre-cleaned sample containers.

At the end of a test run, the mass of liquid collected in each impinger was measured using a scale to within ± 0.5 grams; these masses were used to calculate moisture content of the flue gas. The contents of the impinger train were discarded after the mass is measured.

Bureau Veritas labeled each container with the test number, test location, and test date, and marked the level of liquid on the outside of the container. Immediately after recovery, the sample containers were stored. Bureau Veritas personnel transported the samples to Bureau Veritas' laboratory in Novi, Michigan, for analysis. Figure 4 in the Appendix depicts the USEPA Method 5 sampling train.

4.1.6 Total Hydrocarbons (USEPA Method 25A)

The THC sampling followed USEPA Method 25A, "Determination of Total Gaseous Organic Concentration Using a Flame Ionization Analyzer" procedures. Samples were collected through a stainless steel probe and heated sample line into the analyzer. Bureau Veritas used J.U.M. 109A and/or J.U.M 3-300A flame ionization detector based hydrocarbon analyzers. Figure 5 in the Appendix depicts the USEPA Method 25A sampling train.



A flame ionization detector (FID) determines the average hydrocarbon concentration in part per

million by volume (ppmv) of THC as the calibration gas (i.e., propane). The FID is fueled by 100% hydrogen, which generates a flame with a negligible number of ions. Flue gas is introduced into the FID and enters the flame chamber. The combustion of flue gas generates electrically charged ions. The analyzer applies a polarizing voltage between two electrodes around the flame, producing an electrostatic field. Negatively charged ions, anions, migrate to a collector electrode, while positive charged ions, cations, migrate to a high-voltage electrode. The current between the electrodes is directly proportional to the hydrocarbon concentration in the sample. The flame chamber is depicted at right.

Using the voltage analog signal, measured by the FID, the concentration of volatile organic compounds (VOCs) is recorded by a data acquisition system (DAS). The average concentration of VOCs is reported as the calibration gas (i.e., propane) in equivalent units.



Before testing, the FID analyzers were calibrated by introducing a zero-calibration range gas (<1% of span value) and high-calibration range gas (80-90% span value) to the tip of the sampling probe. The span values were set to 1.5 to 2.5 times the expected concentration (e.g., 0-100 ppmv). Next, a low-calibration range gas (25-35% of span value) and mid-calibration range gas (45-55% of span value) were introduced. The analyzers were considered to be calibrated when the analyzer response was \pm 5% of the calibration gas value.

At the conclusion of a test run a calibration drift test was performed by introducing the zero- and mid-calibration gases to the tip of the sampling probe. The test run data were considered valid if the calibration drift test demonstrated the analyzers responded within $\pm 3\%$ of calibration span from pre-test to post-test calibrations.

4.1.7 Gas Dilution (USEPA Method 205)

A gas dilution system was used to introduce known values of calibration gases into the THC FID gas analyzers. The gas dilution system consisted of calibrated orifices that diluted a high-level calibration gas to within $\pm 2\%$ of predicted values. This gas divider was capable of diluting gases at 80, 60, 50, 30, and 25% increments.

Before the start of testing, the gas divider dilutions were measured to be within $\pm 2\%$ of predicted values. Three sets of dilutions at 80, 60, 50, 30, and 25% of the high-level calibration gas were performed. In addition, a certified mid-level calibration gas was introduced into the analyzer that responded to within $\pm 2\%$ of the expected value. Refer to Appendix A for the Method 205 gas dilution calibration sheets.



4.1.8 Formaldehyde and Methanol (USEPA Method 320)

VOC/HAP emissions were measured in accordance with USEPA Method 320, "Measurements of Vapor Phase Organic and Inorganic Emissions by Extractive Fourier Transform Infrared (FTIR) Spectroscopy." Gaseous samples were withdrawn from the stack and transferred to MKS Instruments MultiGas 2030 FTIR spectrometers for formaldehyde and methanol measurements. Figure 6 in the Appendix depicts the USEPA Method 320 sampling train.

The samples were directed through a heated probe, heated filter and heated transfer line connected to the FTIR. The probes, filters, transfer lines, and FTIRs were maintained at 191° C (375° F) during testing. The formaldehyde and methanol concentrations were measured based on their infrared absorbance compared to reference spectra. The FTIR analyzer scans the sample approximately once per second. A data point consists of the co-addition of 64 scans, with a data point generated every minute.

FTIR quality assurance procedures followed USEPA Method 320. A calibration transfer standard (CTS) was analyzed before and after testing. Acetaldehyde and methanol spiking were performed before and after each test run. Section 3.29 of USEPA Method 320 allows the use of a surrogate analyte for the purposes of analyte spiking. Acetaldehyde was chosen as surrogate to formaldehyde for the following reasons:

- The highest obtainable formaldehyde cylinder is 30 ppm: therefore, the spiked concentration would be 3 ppm (analyte spiking consists of sampling 1 part calibration gas in the presence of 9 parts effluent gas). The formaldehyde concentrations of the sources tested were much higher than 3 ppm.
- Acetaldehyde's physical and chemical properties are similar to those of formaldehyde. Formaldehyde is the C₁ aldehyde (CH₂O); acetaldehyde is the C₂ aldehyde (CH₃CHO).

The analyte spikes were set to a target dilution ratio of 1:10 or less. Valid tests required acetaldehyde and methanol spike recoveries to be within the Method 320 allowance of $\pm 30\%$.

4.2 **Procedures for Obtaining Process Data**

Process data was recorded by Decorative Panels International, Inc. personnel during testing. Refer to Section 2.1 and 2.2 for discussions of process and control device data and Appendix E for the operating parameters recorded during testing.

4.3 Sampling Identification and Custody

Sample identification and chain of custody procedures were applicable to the sampling methods used in this test program. Applicable Chain of Custody procedures followed guidelines outlined within ASTM D4840-99 (Reapproved 2010), "Standard Guide for Sample Chain-of-Custody



Procedures." Detailed sampling and recovery procedures are described in Section 4.0. For each sample collected (i.e. filter) sample identification and custody procedures were completed as follows:

- Containers were sealed with Teflon tape to prevent contamination.
- Containers were labeled with test number, location, and test date.
- The level of fluid was marked on outside of sample containers to identify if leakage had occurred before delivery of the samples to the laboratory.
- Containers were placed in a cooler for storage.

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- Samples were logged using guidelines outlined in ASTM D4840-99 (Reapproved 2010), "Standard Guide for Sample Chain-of-Custody Procedures."
- Samples were delivered to the laboratory.

Chains of custody and laboratory analytical results are included in Appendix F.



5.0 QA/QC Activities

5.1 Pretest QA/QC Activities

Before testing, the sampling equipment was cleaned, inspected, and calibrated according to procedures outlined in the applicable USEPA sampling method and USEPA's "Quality Assurance Handbook for Air Pollution Measurement Systems, Volume and Principles" and, Volume III, "Stationary Source Specific Methods." Refer to Appendix A for inspection and calibration sheets.

5.2 QA/QC Audits

The results of select sampling and equipment QA/QC audits and the acceptable USEPA tolerance are presented in the following sections.

5.2.1 Method 5 QA/QC Audits

The sampling trains described in Section 4.1 were audited for measurement accuracy and data reliability. The following table summarizes the QA/QC audits conducted on each sampling train.

Parameter	Run 1	Run 2	Run 3	Method Requirement	Comment
No. 3 Biofilter Outle	et –				
Average velocity pressure head (in H ₂ O)	1.41	1.39	1.30	>0.05 in H ₂ O [†]	Valid
Sampling train leak check Post–test	0.010 ft ³ for 1 min at 6 in Hg	0.005 ft ³ for 1 min at 5 in Hg	0.005 ft ³ for 1 min at 7 in Hg	<0.020 ft^3 for 1 minute at \ge recorded during test	Valid
Sampling vacuum (in Hg)	0	0	0		

Table 5-1Method 5 Sampling Train QA/QC Audits

† Manometer capable of reading 0 to 10 in H₂O acceptable for measuring differential pressure head above 0.05 in H₂O

5.2.2 Instrument Analyzer QA/QC Audits

The FID and FTIR analyzers met the QA/QC requirements of USEPA Methods 25A and 320. The analyzers were calibrated using USEPA Traceability Protocol calibration gases with an



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uncertainty $\leq 2\%$ of certified value. FID calibration error tests indicated the analyzers were responding to $\pm 5.0\%$ of the cylinder concentration and did not drift more than $\pm 3\%$ before and after each test run. The FTIR analyzers passed all QA/QC procedures included acetaldehyde and methanol spike recoveries within the $\pm 30\%$ allowance.

Refer to Appendix A for the calibration gas certificates and analyzer calibration data and Appendix F for the FTIR calibration data.

5.2.3 Dry-Gas Meter QA/QC Audits

A dry-gas meter was used to sample the flue gas during measurement of moisture content. Table 5-2 summarizes the dry-gas meter (DGM) calibration checks in comparison to the acceptable USEPA tolerance.

Refer to Appendix A for the pre- and post- test DGM calibrations.

Table 5-2DGM Calibration QA/QC Audit

Meter Box	Pre-test DGM Calibration Factor (Y) (dimensionless)	Post-Test DGM Calibration Check Value (Y _{qa}) (dimensionless)	Difference Between Pre- and Post-test DGM Calibrations	Acceptable Tolerance	Comment
7	1.018 (Nov. 25, 2013)	1.015 (Jan. 23, 2014)	0.003	0.05	Valid

5.2.4 Thermocouple QA/QC Audits

Temperature measurements using thermocouples and digital pyrometers were compared to a reference temperature (i.e., ice water bath, boiling water) prior to and after testing to evaluate accuracy of the equipment. The thermocouples and pyrometers measured temperature within $\pm 1.5\%$ of three reference temperatures and, therefore, the equipment met USEPA acceptance criteria. Thermocouple calibration sheets are presented in the Appendix A.

5.3 QA/QC Blanks

Field blanks were analyzed for the constituent of interest. The results of the blanks are presented in Table 5-3. The blank results do not indicate significant contamination occurred in the field. Blank corrections were not applied.



Table 5-3 QA/QC Blanks

Sample Identification	Result (mg)	Comment
M5 Acetone Blank	1.9	Sample volume is 230 milliliters. Reporting limit is 0.5 milligrams. Acetone blank corrections not applied.
M5 Filter Blank	<0.5	Filter blank corrections not applied

5.4 QA/QC Problems

QA/QC problems were not encountered during this test program.



Limitations

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Tables



Table 1

No. 3 Biofilter Formaldehyde, Methanol, and THC Destruction Efficiency Results Decorative Panels International, Inc.

Alpena, Michigan Bureau Veritas Project No. 11014-000010.00 Sampling Date: January 9, 2014

Parameter		Date	1/9/2014	1/9/2014	1/9/2014	
		Units	Run 1	Run 2	Run 3	Average
Sampling Start Time		hh:mm	10:45		18:21	
Duration		min	60	60	60	60
Average	Biobed temperature	°F	82	82	82	82
Inlet						
	Gas Stream Volumetric Flowrate	scfm	37,110	38,693	39,432	38.412
1		1	1		1	1
	Formaldehyde Concentration	ppmv, as CH ₂ O	19.1	17.5	19.7	18.8
	Formaldehyde Mass Emission Rate	lb/hr, as CH ₂ O	3.3	3.2	3.6	3.4
]	Methanol Concentration	ppmv, CH₃OH	24.7	22.9	23.6	23.7
	Methanol Mass Emission Rate	lb/hr, as CH3OH	4.6	4.4	4.6	4.5
ļ						
	THC Concentration	ppmv, as propane	45.0	53.2	50.9	49.7
	THC Concentration	ppmv, as carbon	135	160	153	149
1	THC Mass Emission Rate	lb/hr, as propane	11	14	14	13
	THC Mass Emission Rate	lb/hr, as carbon	9	12		11
Outlet						
1	Gas Stream Volumetric Flowrate	scfm	57,176	57,138	54,999	56.438
ļ						0.0
	Formaldehyde Concentration	ppmv, as CH ₂ O	0.3	0.3	0.4	0.5
]	Formaldehyde Mass Emission Rate	Ib/hr, as CH ₂ O	0.1	0.1	0.1	0.1
						0.0
	Methanol Concentration	ppmv, CH ₃ OH	1.0	0.6	0.9	0.8
1	Methanol Mass Emission Rate	lb/hr, as CH ₃ OH	0.3	0.2	0.3	0.2.
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	THC Concentration	ppmv, as propane	1.2	8.5	9.4	ð.4
	THC Concentration	ppmv, as carbon	21.0	25.5	28.1	23.1
	THC Mass Emission Rate	ib/nr, as propane	2.0	5.5	5.1	3.3
ļ	THC Mass Emission Rate	lb/hr, as carbon	2.3	2.7	3.0	2.7
Formal	hehyde Destruction Efficiency Results	%	97.6	97.5		97.5
Methan	ol Destruction Efficiency Results	<u>%</u>	93.8	96.1	94.4	94.7
THC D	estruction Efficiency Results	%	75.4	76.4	73.4	75.0



Table 2 - No. 3 Biofilter Particulate Matter Results							
Facility		inc.					
Source Designation Test Date		No. 3 B January 9, 2014	January 9, 2014	January 9, 2014			
Meter/Nozzle Information		Run 1	Run 2	Run 3	Average		
Meter Temperature, T _m	°F	53	62	61	59		
Meter Pressure, Pm	in Hg	30.62	30,64	30.62	30.63		
Measured Sample Volume, Vm	ft3	41,27	42.54	41.58	41.80		
Sample Volume, V.,	std A'	44.29	44,85	43.90	44.34		
Samule Volume, V.	stđ m ³	1.25	1.27	1.24	1.20		
Condensate Volume, V.,	sid ft ³	2.02	2.32	1.98	2.1		
Gas Density, o.	sid lb/fl ³	0.0736	0.0735	0 0736	0.0736		
Total weight of sampled gas	Ъ	3.410	3,464	3.324	3,400		
Nozzle Size, A	A ²	0.0002032	0.0002032	0.0002032	0.0002032		
Isokinetic Variation, I	%	95	97	98	97		
Stack Data							
Average Stack Temperature, T,	°F	86	88	87	87		
Molecular Weight Stack Gas-dry, M _J	lb/lb-mole	28,84	28,83	28.84	28.84		
Molecular Weight Stack Gas-wet, M,	Ib/Ib-mole	28.36	28.30	28.37	28.34		
Stack Gas Specific Gravity, G,		0.98	0.98	0.98	0.98		
Percent Moisture, B _{ws}	%	4.37	4.91	4,32	4.53		
Water Vapor Volume (fraction)		0.044	0.049	0.043	0.045		
Pressure, P,	in Hg	30.69	30,69	30.69	30.69		
Average Stack Velocity, V,	fl/sec	67.08	67,21	64.62	66.30		
Area of Stack	η²	14.33	14.33	14.33	14,33		
Exhausi Gas Flowrate							
Flowrate	ft ³ /min, actual	57,657	57,76 7	55,546	56,990		
Flowrate	A ³ /min, standard wet	57,176	57,138	54,999	56,438		
Flowrate	ft ³ /min, standard dry	54,679	54,333	52,625	53,879		
Flowrate	m³/min, standard dry	1,548	1,539	1,490	1,526		
Collected Mass							
Acetone Wash	mg	45	12	79	45		
Filter	mg	<0.5	<0,5	0.80	0.6		
Total Filterable Particulate Matter (FPM)	mg	46	13	80	46		
Concentration							
Particulate Matter (FPM)	mg/dscf	1.0	0.28	1.8	1.0		
Particulate Matter (FPM)	grain/dscf	0.016	0.0043	0.028	0.016		
Particulate Matter (FPM)	1b/1,000 lb	0.030	0.0082	0.054	0.031		
Mass Emission Rate							
Particulate Matter (FPM)	lb/hr	7.4	2.0	13	7.4		

Figures

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Graphs





